

**PHYTOCHEMICAL CHARACTERIZATION OF CINNAMOMUM
ZEYLANICUM (CINNAMON) BARK**

BY

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HND/23/SLT/FT/0844**

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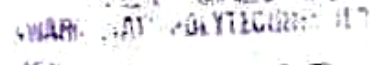
CERTIFICATION

This is to certify that this project work presented by DAUDA SHUKURAH ARAMIDE with Matric No. HND/23/SLT/FT/0844, has been read, approved and submitted to the Department of Science Laboratory Technology (Biochemistry Unit), Institute of Applied Sciences, Kwara State Polytechnic, Ilorin.


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DEDICATION

This project is dedicated to Almighty God, the most beneficent, the most merciful, my source of inspiration, who has showered me with His bountiful mercy. And to my lovely parents, **MR. & MRS. DAUDA**

ACKNOWLEDGEMENTS

My immeasurable gratitude to Almighty Allah, who has sustained me, strengthen me, guide me and spare my life till this present level of education, I have attained. I also Acknowledge our noble prophet Mohammed(S.A.W)his household and his companionship till the day of resurrection.

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ABSTRACT

13.85g of dry cinnamon powder was extracted with 300ml methanol to yield 37% of crude extract using the soxhlet method. The phytochemical analysis using standard methods revealed the presence of alkaloids, triterpenes, lactones, and phytosterols. There was an absence of saponins and glycosides.

CHAPTER ONE:

INTRODUCTION AND LITERATURE REVIEW

1.1 INTRODUCTION

The bark from different cinnamon species holds significant importance as a widely used spice globally. Not just in culinary practices but also in both traditional and modern medicinal applications. The cinnamon genus comprises around 250 identified species, and trees distributed across the world (Sangal 2011).

The primary utilization of cinnamon is in the fragrance and essence industries, leveraging on its aromatic qualities that can enhance various food items, perfumes and medicinal products. Cinnamon's essential components namely *cis*-cinnamaldehyde and *trans*-cinnamaldehyde (*cin*), found in its essential oil play a crucial role in both its fragrance and the diverse biological activities associated with cinnamon.

Research on *Cinnamom osmophloeum* (*C. osmophloeum*) revealed that the essential oil extracted from cinnamon possesses a notable concentration of *cin*. As a result, *C. osmophloeum* is employed as a substitute spice for *C. cassia*. (*E*)-cinnamaldehyde, a significant component in the essential oil is derived from *C.*

zeylanicum exhibit antityrosinase activity with cinnamaldehyde identified as the primary compound responsible for this particular activity. (Chao 2013)

Procyanidins and catechins are present in cinnamon bark. The procyanidins in cinnamon bark encompass both A-type and B-type linkages. The procyanidins obtained from both cinnamon and berries exhibit antioxidant properties. (Pang 2008)

1.2 SCIENTIFIC CLASSIFICATION OF CINNAMON

Kingdom: Plantae

Division: Tracheophyta

Class: Magnoliopsida

Order: Laurales

Family: Lauraceae

Genus: Cinnamomum

Species: Zeylanicum

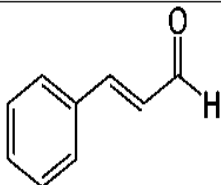
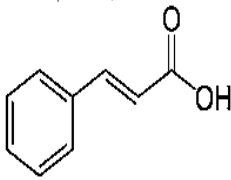
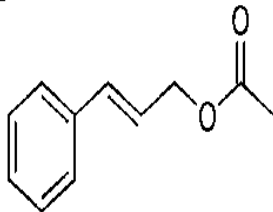
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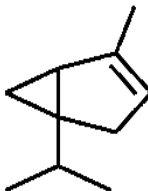
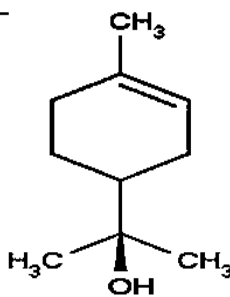
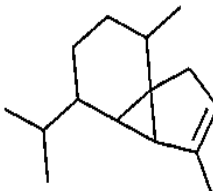
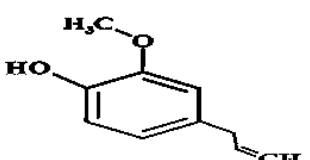
1.3 CHEMICAL CONSTITUENTS

Cinnamon comprises diverse resinous compounds, including cinnamaldehyde, cinnamate, cinnamic acid, and various essential oils (Senanayake, Lee & Wills 1978) The occurrence of an extensive array of essential oils, including trans-cinnamaldehyde, cinnamyl acetate, eugenol, L-borneol, caryophyllene oxide, β -caryophyllene, L-bornyl acetate, E-nerolidol, α -cubebene, α -terpineol, terpinolene, and α -thujene, has been documented.(Tung et al. 2008)

The chemical structures of some important constituents of cinnamon are shown in Table,

Table 1: Chemical structures of some important constituents of cinnamon

Name	Structure
Cinnamaldehyde	
Cinnamic acid	
Cinnamyl Acetate	

α-Thujene	
α-terpineol	
α-cubebene	
Eugenol	

1.4 TRADITIONAL USES:

Besides its role as a spice and flavoring agent, cinnamon finds application in flavoring chewing gums, thanks to its mouth-refreshing effects and capability to combat bad breath. Cinnamon also has the potential to enhance colon health, thereby lowering the risk of colon cancer. (Wondiak 2010)

Cinnamon functions as a coagulant, preventing bleeding. Cinnamon additionally enhances blood circulation in the uterus and promotes tissue regeneration. While

playing a crucial role as a spice, this plant's essential oils and other constituents exhibit significant activities, including antimicrobial, antifungal, antioxidant, and antidiabetic properties. (Gende 2008)

Cinnamon has been employed as an anti-inflammatory, antitermitic, nematocidal, mosquito larvicidal, insecticidal, antimycotic, and anticancer agent. Traditionally, cinnamon has been utilized as tooth powder to address toothaches, dental problems, oral microbiota, and bad breath (Aneja 2009).

1.5 ANTI-INFLAMMATORY ACTIVITIES:

Cinnamon has been found to possess anti-inflammatory properties in numerous studies focusing on medicinal plants and their constituents. Several studies have documented the anti-inflammatory properties exhibited by cinnamon and its essential oils. As of now, numerous flavonoid compounds such as gossypin, gnaphalin, hesperidin, hibifolin, hypolaetin, oroxindin, and quercetin have been identified and exhibit anti-inflammatory properties. (Garia-lafuente 2009)

A recent research study revealed that 2-hydroxycinnamaldehyde, derived from *C. cassia* bark, demonstrated inhibitory effects on nitric oxide production by suppressing the activation of nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B). This suggests its potential as an anti-inflammatory agent. The ethanolic extract from *C. cassia* demonstrated notable anti-

inflammatory effects by decreasing the activation of Src/spleen-tyrosine-kinase- (Src/Syk-) mediated NF- κ B. Compounds found in *C. ramulus* exhibit anti-inflammatory properties by inhibiting the expression of inducible nitric oxide synthesis (iNOS), cyclooxygenase-2 (COX-2), and nitric oxide (NO) production in the central nervous system (CNS). This suggests that *C. ramulus* has the potential to be utilized for therapeutic treatment or prevention of neurodegenerative diseases associated with inflammation. Moreover, the cinnamon's aqueous extract reduces the levels of tumor necrosis factor- α in the serum induced by lipopolysaccharide. Alzheimer's disease results in a significant decrease in 56 kDa. An oligomer, leading to a reduction in plaques and enhancement of cognitive performance in transgenic mouse models (Frydman-Marom, 2011).

Another research found that the water-based extract from *C. zeylanicum* has the potential to diminish tau aggregation and filament formation, key characteristics of Alzheimer's disease. Additionally, the extract promotes the complete fragmentation of recombinant tau filaments and induces significant alterations in the morphology of paired helical filaments derived from Alzheimer's disease brains. (Peterson, 2009)

1.6 ANTICANCER ACTIVITY

The HPLC-derived aqueous extract and cinnamon fraction rich in procyanidins effectively hinder the kinase activity of vascular endothelial growth factor subtype 2 (VEGFR2), consequently impeding angiogenesis associated with cancer. Findings suggest the potential utility of cinnamon in cancer prevention. Cinnamaldehydes were synthesized and assessed for their inhibitory effects on angiogenesis. Jeong and colleagues discovered that CB403, a compound derived from 2-hydroxycinnamaldehyde found in cinnamaldehyde, demonstrated the ability to hinder tumor growth. The findings from both animal-based and cell culture studies suggest the potential use of cinnamon as an anticancer agent due to the antitumor and growth-inhibitory properties of CB403. (Jeong 2003)

According to Cabello and colleagues (2009) the cinnamic aldehyde hinders NF- κ B activity and suppresses the production of interleukin-8 (IL-8) induced by tumor necrosis factor alpha (TNF α -) in A375 cells. This restraint further reinforces the overlooked potential of cinnamic acid in its role as a potential anticancer agent.

1.7 ANTIOXIDANT ACTIVITY:

Antioxidants found in food are essential for human health, serving as protective agents. Apart from their crucial role, antioxidants are integral additives in fats and oils. In the food processing industry, they are utilized to extend the shelf life and

prevent spoilage. Spices and medicinal plants are gaining quick recognition as valuable sources of antioxidants with potential health benefits against various diseases. [49] Antioxidants play a crucial role in sustaining human and animal life by combating free radicals and addressing damage associated with metabolic diseases and age-related syndromes. (Halliwell, 2011)

1.8 AIMS & OBJECTIVE

To identify and qualitatively screen the major phytochemical groups present in the bark, such as alkaloids, glycosides, tannins, flavonoids, steroids, saponins, essential oils, and phenolic compounds.

To assess the presence of bioactive compounds that contributes to antioxidant, antimicrobial, anti-inflammatory, and other medicinal properties of cinnamon bark.

To quantify key phytochemicals like total phenols, flavonoids, tannins, and saponins to understand their abundance and potential health benefits.

To evaluate the antioxidant activity linked to these phytochemicals, which may help in scavenging free radicals and preventing diseases such as cancer.

To provide scientific evidence supporting the use of cinnamon bark as a natural source for pharmaceuticals, nutraceuticals, food additives, and cosmetics.

To investigate the safety profile by screening for toxic compounds and heavy metals to ensure the extracts suitability for medicinal and dietary use.

To explore the chemical composition of essential oils in the bark and the biological effect, including potential tissue- protective actions.

CHAPTER TWO:

MATERIAL AND METHODS

2.1 COLLECTION OF PLANT MATERIAL

The fresh sample of plant material (cinnamomum Zeylanicum bark) was purchased at Oja Oba in Ilorin.

2.2 APPARATUS AND GLASSWARE

Here's a list of common apparatus and Glassware used in the extraction and phytochemical testing of cinnamon bark.

2.2.1 Apparatus

- * Soxhlet extractor
- * Rotary evaporator
- * Heating mantle
- * Water bath
- * Spectrophotometer (for quantitative analysis)
- * Hot Plate
- * Mortar and Pestle (for grinding)

2.2.1 Glassware

*Round-bottom flask

* Conical flasks

* Beakers

* Measuring cylinders

* Pipettes

* Test tubes

* Petri dishes

2.3 CHEMICALS AND REAGENTS

All chemicals and reagents utilized in the study were of analytical grade.

Ethyl alcohol used for extraction (about 4 liters), 70% v/v (70ml absolute ethanol: 30ml distilled water).

The reagents used for detecting various phytochemical groups include Wagner's, Dragendorff's, Hager's reagent (for alkaloid detection), Benedict's, Fehling's, Molisch's reagent (for Glycosides) vanillin (for gallic acid), hydrochloric acid reagent (for tannins/phenols). Wilson's and Alkaline reagents (for phlobatannins) lead acetate, gelatin, FeCl_3 reagents (for flavonoids) Absolute alcohol (for gum), Distilled water (for resins), and Biuret reagent (for proteins) (Audu S.A et al, 2007).

2.4 EXTRACTION OF PLANT MATERIAL

The plant material of *cinnamomum zeylanicum* was pulverized using a high powered multifunctional kitchen blender SAMSUNG (Model No:2022L) with 500w and 32000Rp, made in Japan.

13.85g of the sample was packed into a cellulose thimble and placed in a 1L beaker. 300ml of methanol solvent was measured and transferred into the beaker to cover the sample in the thimble. A magnetic bar was placed at the bottom of the beaker. The beaker and its content were placed on a magnetic stirrer temperature regulated hot plate. (Pandey et al., 2014).

The soxhlet extraction was done for 2hrs. the colored extract solution was removed and another 300ml of fresh methanol was added, and extraction process repeated until the sample was exhaustively extracted. All the extraction was pooled together and transferred into a 1L round bottom flask the extract solution was distilled to remove the methanol solvent. The concentrated extract was subsequently transferred into a beaker and placed in a water bath, heating beaker and heating was done until all solvent was almost completely evaporated. The beaker and its content were left cool at ambient temperature until it dried.

The weight of crude extract obtained was determined, from which the extract yield was calculated, the crude methanol extract of 5.21g was obtained and kept in the laboratory at ambient temperature for further analysis.

2.5 PHYTOCHEMICALS TEST OF EXTRACT

The extract was tested for the presence of bioactive compound by using the following standard methods.

2.5.1 Test for Steroids

2.5.1.1 Salkowski Test: the extract was mixed with 2ml of chloroform and concentrated Sulphuric acid was added along the sides of the test tube and on standing yields red color. (Trifan et al., 2021).

2.5.1.2 Lieberman Burchard test: The extract was mixed with 2ml of chloroform and few drops of acetic anhydride and 1ml of concentrated sulphuric acid from the sides gives reddish ring at the junction of 2 layers. (sadasivam s et al., 2005).

2.5.2 Test for Triterpenes

2.5.2.1 Salkowski Test: he extract was mixed with 2ml of chloroform and a few drops of acetic acid and 1ml of concentrated sulphuric acid given deep red at the junction of 2 layers.

2.5.3 Test for Alkaloids

The extract was dilute with ammonia and then extracted with chloroform solution to this dilute hydrochloric acid was added. The acid was used for chemical test of alkaloids.

2.5.3.1 Mayer's test (potassium mercuric iodide): The acid layer with few drops of Mayer's reagent given a creamy white precipitate.

2.5.3.2 Wagner's test (sodium of iodine in potassium iodide)

The extract with a few drops of Wagner's reagent gives reddish brown colored precipitate (Prassas et al., 2008).

2.5.3.3 Hager's test: (solution of iodine in potassium iodide).

The extract with Hager's reagent gives yellow precipitate

2.5.3.4 Dragendoff's test (solution of potassium Bismuth iodide).

The extract with a few drops of dragendoff's reagent gives a reddish brown precipitate

2.5.4 Test for Tannin: Ferric chloride test

The extract with 10% ferric chloride solution gives brownish green colour.

2.5.5 Test for Lactones: Legals test:

The extract with the mixture of sodium nitroprusside and pyridine and treated with methanol alkali gives deep red colour.

2.5.6 Test for Flavonoid Lead acetate test:

The extract was mixed with few drop of 10% lead acetate gives a yellow precipitate. (Raanan N. et al., 2006).

2.5.7Test for Diterpene.

Copper acetate test: The extract was mixed with copper acetate solution gives green colour.

2.5.8Test for Glycosides

Sodium hydroxide reagent: A small amount of alcoholic extract was dissolved in water and sodium hydroxide solution was added gives a yellow colour (Vakilwala et al 2017).

2.5.9Test of saponin

Flax extract was mixed with 2ml of distilled water in a test tube the mixture was shaking vigorously and observed for the formation of persistant confirms the and observe from that presence of saponin (Rainsfird et al., 2018)

CHAPTER THREE

3.0 RESULT AND DISCUSSION

3.1 RESULT

The result of qualitative phytochemical characterization of cinnamon bark is presented in the table below.

TABLE 3.1 RESULT OF PHYTOCHEMICAL TEST

PHTOCHEMICAL	RESULTS
Alkaloids	+
Triterpene	+
Tannins	-
Diterpenes	+
Sterols	+
Glycosides	-
Flavonoids	+
Lactones	+
Saponins	-

NOTES: + = PRESENT - = ABSENT

3.1.1 Percentage calculation of extract yield

The percentage crude extract yield of phytochemical characterization of (Cinnamon) is calculated as followed.

$$\% \text{ Extract yield} = \frac{\text{Weights of crude extract} \times 100}{\text{Weight of cinnamon sample}}$$

Given value

Weight of crude extract=5.21g

Weight of cinnamon sample=13.85g

$$\begin{aligned} \% \text{ Extract yield} &= \text{weight} = \frac{5.21\text{g}}{13.85\text{g}} \times 100 \\ &= 37\% \end{aligned}$$

Therefore, the percentage of extract yield = 37%

DISCUSSION

The dark brown sticky paste crude extract gave a percentage yield of 37%.

The result in (Table 3.1) show the phytochemical properties of cinnamon bark which involves the cinnamon bark chemical compound that contributes to its medicinal properties. These compounds that includes alkaloids steroids, tannins, flavonoids, lactones, diterpenes, glycosides, saponnins and titerpenes which are

known for their antioxidant, anti-inflammatory, anticancer, anti-bacterial, anti-fungal and antimicrobial activity.

The phytochemical analysis conducted allows us to identify and qualify these bioactive compounds, providing valuable insights into the potential health benefits of the study. (Vakulwala 2017)

CONCLUSION

Cinnamon has been utilized as a spice in daily life without observable side effects. Numerous reports have delved into the diverse properties of cinnamon, including its bark, essential oils, bark powder, phenolic compounds, Flavonoids, and isolated components. Each of these properties plays a pivotal role in advancing human health. Antioxidant and antimicrobial activities may result from direct action on oxidants or microbes, while anti-inflammatory, anticancer, and antidiabetic activities occur indirectly through receptor-mediated mechanisms. Extensive exploitation has unveiled significant health benefits across various types of cinnamon. Further investigations are required to provide additional uses of this spice against cancer and inflammatory, cardio protection, and neurological disorder.

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