

**COORDINATION CHEMISTRY OF SYNTHESIS,
CHARACTERIZATION AND BIOLOGICAL ACTIVITY OF
CHLOROQUINE AND SELECTED D-BLOCK METALS.**

A REPORT SUBMITTED

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CERTIFICATION

This is to certify that this project work was carried out **AMUSAT TOHEEB**
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DEDICATION

This research work is dedicated to the Almighty God, the creator of heaven and earth. I also dedicate it to my beloved parents Mr. and Mrs. **AMUSAT** for their care and support towards me right from my childhood

ACKNOWLEDGEMENT

My sincere heart of gratitude goes to the Almighty Allah who gave me knowledge and strength for the successful completion of this HIGHER National Diploma program.

This project work would not have been possible without the cooperation of a very large number of people. My greatest debt in this regard however, is to my supervisor, in person of Mr. ABDULGANIYU B.B whose patient criticisms and comments played a big role towards the completion of this project work. I am extremely grateful to him.

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My heartfelt appreciation also goes to all my family members.

This journey has been one of determination, resilience, and unwavering self-reliance. I take pride in acknowledging that I sponsored myself throughout my academic pursuit, without any external financial assistance.

Every milestone was achieved through the consistent efforts, hard work, and support of my family.

This success story is a testament to what self-belief and dedication can accomplish.

ABSTRACT

Some Chloroquine complexes of Ni[ii], Cu[ii] and Iron[ii] were prepared from the reaction of chloroquine and metal salts.

The complexes were characterized using physical properties like melting point and solubility test.

LIST OF ABBREVIATIONS

Ni	Nickel
Zn	Zinc
Fe	Iron
S	Soluble
Ns	Not Soluble
Ss	Slightly Soluble
Uv	Ultraviolet
IR	InfraRed
g/mol	Gram per mole.

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CHAPTER ONE

1.1 INTRODUCTION

Chloroquine is a medication primarily used to prevent and treat malaria in areas where malaria remains sensitive to its effects.[1] Certain types of malaria, resistant strains, and complicated cases typically require different or additional medication.[1] Chloroquine is also occasionally used for amebiasis that is occurring outside the intestines, rheumatoid arthritis, and lupus erythematosus.[1] While it has not been formally studied in pregnancy, it appears safe.[1][2] It was studied to treat COVID-19 early in the pandemic, but these studies were largely halted in the summer of 2020, and the NIH does not recommend its use for this purpose.[3] It is taken by mouth.[1]

Common side effects include muscle problems, loss of appetite, diarrhea, and skin rash.[1] Serious side effects include problems with vision, muscle damage, seizures, and low blood cell levels.[1][4] Chloroquine is a member of the drug class 4-aminoquinoline.[1] As an antimalarial, it works against the asexual form of the malaria parasite in the stage of its life cycle within the red blood cell.[1] How it works in rheumatoid arthritis and lupus erythematosus is unclear.[1]

Chloroquine was discovered in 1934 by Hans Andersag.[5][6].It is on the World Health Organization's List of Essential Medicines.[7]It is available as a generic medication.[1]

Chloroquine has been used in the treatment and prevention of malaria from *Plasmodium vivax*, *P. ovale*, and *P. malariae*. It is generally not used for *Plasmodium falciparum* as there is widespread resistance to it.[9][10]

Chloroquine has been extensively used in mass drug administrations, which may have contributed to the emergence and spread of resistance. It is recommended to check if chloroquine is still effective in the region prior to using it.[11]In areas where resistance is present, other antimalarials, such as mefloquine or atovaquone, may be used instead. The Centers for Disease Control and Prevention recommend against treatment of malaria with chloroquine alone due to more effective combinations.[12]

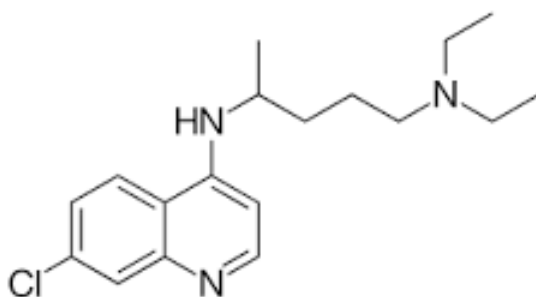


Fig. 1 Structure of Chloroquine.

1.2 Rheumatic disease

As it mildly suppresses the immune system, chloroquine is used in some autoimmune disorders, such as rheumatoid arthritis and has an off-label indication for lupus erythematosus.[1]

1.3 Side effects

Side effects include blurred vision, nausea, vomiting, abdominal cramps, headache, diarrhea, swelling legs/ankles, shortness of breath, pale lips/nails/skin, muscle weakness, easy bruising/bleeding, hearing and mental problems.[14][15]

- Unwanted/uncontrolled movements (including tongue and face twitching, diskinesia, and dystonia)[14][16]
- Deafness or tinnitus[14]
- Nausea, vomiting, diarrhea, abdominal cramps[15]
- Headache[14]
- Mental/mood changes (such as confusion, personality changes, unusual thoughts/behavior, depression, feeling being watched, hallucinating)[14][15]
- Signs of serious infection (such as high fever, severe chills, persistent sore throat)[14]
- Skin itchiness, skin color changes, hair loss, and skin rashes[15][17]

- Chloroquine-induced itching is very common among black Africans (70%), but much less common in other races. It increases with age, and is so severe as to stop compliance with drug therapy. It is increased during malaria fever; its severity is correlated to the malaria parasite load in blood. Some evidence indicates it has a genetic basis and is related to chloroquine action with opiate receptors centrally or peripherally.[18]
- Triggering of a severe psoriasis attack in those with psoriasis[16]
- Unpleasant metallic taste
 - This could be avoided by "taste-masked and controlled release" formulations such as multiple emulsions.[19]
- Chloroquine retinopathy (irreversible retinal damage)[16]
- Electrocardiographic changes[20]
 - This manifests itself as either conduction disturbances (bundle-branch block, atrioventricular block) or cardiomyopathy — often with hypertrophy, restrictive physiology, and congestive heart failure. The changes may be irreversible. Only two cases have been reported requiring heart transplantation, suggesting this particular risk is very low. Electron microscopies of cardiac biopsies show pathognomonic cytoplasmic inclusion bodies.

- Pancytopenia, aplastic anemia, reversible agranulocytosis, low blood platelets, neutropenia[16]
- Worsening of the condition for those with porphyria[16]

1.4 Pregnancy

Chloroquine has not been shown to have any harmful effects on the fetus when used in the recommended doses for malarial prophylaxis.[21] Small amounts of chloroquine are excreted in the breast milk of lactating women. However, this drug can be safely prescribed to infants, the effects are not harmful. Studies with mice show that radioactively tagged chloroquine passed through the placenta rapidly and accumulated in the fetal eyes which remained present five months after the drug was cleared from the rest of the body.[16][22] Women who are pregnant or planning on getting pregnant are still advised against traveling to malaria-risk regions.[21]

1.5 Elderly

There is not enough evidence to determine whether chloroquine is safe to be given to people aged 65 and older. Since it is cleared by the kidneys, toxicity should be monitored carefully in people with poor kidney functions, as is more likely to be the case in the elderly.[16]

1.6 Drug interactions

Chloroquine has a number of drug–drug interactions that might be of clinical concern

- Ampicillin – levels may be reduced by chloroquine;[16]
- Antacids – may reduce absorption of chloroquine;[16]
- Cimetidine – may inhibit metabolism of chloroquine; increasing levels of chloroquine in the body;[16]
- Cyclosporine – levels may be increased by chloroquine;[16] and
- Mefloquine – may increase risk of convulsions.[16]

1.7 Overdose

Chloroquine, in overdose, has a risk of death of about 20%.[23] It is rapidly absorbed from the gut with an onset of symptoms generally within an hour.[24] Symptoms of overdose may include sleepiness, vision changes, seizures, stopping of breathing, and heart problems such as ventricular fibrillation and low blood pressure.[23][24] Low blood potassium may also occur.[23]

While the usual dose of chloroquine used in treatment is 10 mg/kg, toxicity begins to occur at 20 mg/kg, and death may occur at 30 mg/kg.[23] In children as little as a single tablet can be fatal.[24][16]

Treatment recommendations include early mechanical ventilation, cardiac monitoring, and activated charcoal.[23] Intravenous fluids and vasopressors may be required with epinephrine being the vasopressor of choice.[23] Seizures may be treated with benzodiazepines.[23] Intravenous potassium chloride may be required, however this may result in high blood potassium later in the course of the disease.[23] Dialysis has not been found to be useful.[23]

Absorption of chloroquine is rapid and primarily happens in the gastrointestinal tract.[25] It is widely distributed in body tissues.[26] Protein binding in plasma ranges from 46% to 79%.[27] Its metabolism is partially hepatic, giving rise to its main metabolite, desethylchloroquine.[28] Its excretion is $\geq 50\%$ as unchanged drug in urine, where acidification of urine increases its elimination. It has a very high volume of distribution, as it diffuses into the body's adipose tissue.

Accumulation of the drug may result in deposits that can lead to blurred vision and blindness.[29] It and related quinines have been associated with cases of retinal toxicity, particularly when provided at higher doses for longer times. With long-term doses, routine visits to an ophthalmologist are recommended.

Chloroquine is also a lysosomotropic agent, meaning it accumulates preferentially in the lysosomes of cells in the body. The pKa for the quinoline nitrogen of chloroquine is 8.5, meaning it is about 10% deprotonated at physiological pH (per

the Henderson-Hasselbalch equation). This decreases to about 0.2% at a lysosomal pH of 4.6. Because the deprotonated form is more membrane-permeable than the protonated form, a quantitative "trapping" of the compound in lysosomes results.

1.8 Malaria

The lysosomotropic character of chloroquine is believed to account for much of its antimalarial activity; the drug concentrates in the acidic food vacuole of the parasite and interferes with essential processes. Its lysosomotropic properties further allow for its use for *in vitro* experiments pertaining to intracellular lipid related diseases,[30][31] autophagy, and apoptosis.[32]

Inside red blood cells, the malarial parasite, which is then in its asexual lifecycle stage, must degrade hemoglobin to acquire essential amino acids, which the parasite requires to construct its own protein and for energy metabolism. Digestion is carried out in a vacuole of the parasitic cell.[*citation needed*]

Hemoglobin is composed of a protein unit (digested by the parasite) and a heme unit (not used by the parasite). During this process, the parasite releases the toxic and soluble molecule heme. The heme moiety consists of a porphyrin ring called Fe(II)-protoporphyrin IX (FP). To avoid destruction by this molecule, the parasite

biocrystallizes heme to form hemozoin, a nontoxic molecule. Hemozoin collects in the digestive vacuole as insoluble crystals.

Chloroquine enters the red blood cell by simple diffusion, inhibiting the parasite cell and digestive vacuole. Chloroquine (CQ) then becomes protonated (to CQ²⁺), as the digestive vacuole is known to be acidic (pH 4.7); chloroquine then cannot leave by diffusion. Chloroquine caps hemozoin molecules to prevent further biocrystallization of heme, thus leading to heme buildup. Chloroquine binds to heme (or FP) to form the FP-chloroquine complex; this complex is highly toxic to the cell and disrupts membrane function. Action of the toxic FP-chloroquine and FP results in cell lysis and ultimately parasite cell autodigestion.[33] Parasites that do not form hemozoin are therefore resistant to chloroquine.[34]

1.9 Resistance in malaria

Since the first documentation of *P. falciparum* chloroquine resistance in the 1950s, resistant strains have appeared throughout East and West Africa, Southeast Asia, and South America. The effectiveness of chloroquine against *P. falciparum* has declined as resistant strains of the parasite evolved.

Resistant parasites are able to rapidly remove chloroquine from the digestive vacuole using a transmembrane pump. Chloroquine-resistant parasites pump

chloroquine out at 40 times the rate of chloroquine-sensitive parasites; the pump is coded by the *P. falciparum* chloroquine resistance transporter (*PfCRT*) gene.[35] The natural function of the chloroquine pump is to transport peptides: mutations to the pump that allow it to pump chloroquine out impairs its function as a peptide pump and comes at a cost to the parasite, making it less fit.[36]

Resistant parasites also frequently have mutation in the ABC transporter *P. falciparum* multidrug resistance (*PfMDR1*) gene, although these mutations are thought to be of secondary importance compared to *PfCRT*. An altered chloroquine-transporter protein, *CG2* has been associated with chloroquine resistance, but other mechanisms of resistance also appear to be involved.[37]

Verapamil, a Ca²⁺ channel blocker, has been found to restore both the chloroquine concentration ability and sensitivity to this drug. Other agents which have been shown to reverse chloroquine resistance in malaria are chlorpheniramine, gefitinib, imatinib, tariquidar and zosuquidar.[38]

As of 2014 chloroquine is still effective against poultry malaria in Thailand. Sohsuebngarm et al. 2014 test *P. gallinaceum* at Chulalongkorn University and find the parasite is not resistant.[39]; Sertraline, fluoxetine and paroxetine reverse chloroquine resistance, making resistant biotypes susceptible if used in a cotreatment.[40]

1.10 Antiviral

Chloroquine has antiviral effects against some viruses.[41] It increases late endosomal and lysosomal pH, resulting in impaired release of the virus from the endosome or lysosome — release of the virus requires a low pH. The virus is therefore unable to release its genetic material into the cell and replicate.[42][43]

Chloroquine also seems to act as a zinc ionophore that allows extracellular zinc to enter the cell and inhibit viral RNA-dependent RNA polymerase.[44][45]

1.11 Chemical synthesis

The first synthesis of chloroquine was disclosed in a patent filed by IG Farben in 1937.[44] In the final step, 4,7-dichloroquinoline was reacted with 1-diethylamino-4-aminopentane. By 1949, chloroquine manufacturing processes had been

1.12 Formulations

Chloroquine comes in tablet form as the phosphate, sulfate, and hydrochloride salts. Chloroquine is usually dispensed as the phosphate.[36]

1.13 Names

Brand names include Chloroquine FNA, Resochin, Dawaquin, and Lariago.[37]

Chloroquine, in various chemical forms, is used to treat and control surface growth of anemones and algae, and many protozoan infections in aquariums,[38] e.g. the fish parasite *Amyloodinium ocellatum*. [39] It is also used in poultry malaria.[39]:

Chloroquine was proposed as a treatment for SARS, with *in vitro* tests inhibiting the severe acute respiratory syndrome coronavirus (SARS-CoV).[30][31] In October 2004, a published report stated that chloroquine acts as an effective inhibitor of the replication of SARS-CoV in vitro.[60] In August 2005, a peer-reviewed study confirmed and expanded upon the results.[22]

Chloroquine was being considered in 2003, in pre-clinical models as a potential agent against chikungunya fever.[23]

Chloroquine and hydroxychloroquine are anti-malarial medications also used against some auto-immune diseases.[64] Chloroquine, along with hydroxychloroquine, was an early experimental treatment for COVID-19.[45] Neither drug has been useful to prevent or treat SARS-CoV-2 infection.[46] Administration of chloroquine or hydroxychloroquine to

COVID-19 patients has been associated with increased mortality and adverse effects, such as QT prolongation.[42][43] Researchers estimate that off-label use of hydroxychloroquine in hospitals during the first phase of the pandemic caused 17,000 deaths worldwide.[44] The widespread administration of chloroquine or hydroxychloroquine, either as monotherapies or in conjunction with azithromycin, has been associated with deleterious outcomes, including QT interval prolongation. As of 2024, scientific evidence does not substantiate the efficacy of hydroxychloroquine, with or without the addition of azithromycin, in the therapeutic management of COVID-19.[42]

Cleavage of the SARS-CoV-2 S2 spike protein required for viral entry into cells can be accomplished by proteases TMPRSS2 located on the cell membrane, or by cathepsins (primarily cathepsin L) in endolysosomes.[45] Hydroxychloroquine inhibits the action of cathepsin L in endolysosomes, but because cathepsin L cleavage is minor compared to TMPRSS2 cleavage, hydroxychloroquine does little to inhibit SARS-CoV-2 infection.[45]

Several countries initially used chloroquine or hydroxychloroquine for treatment of persons hospitalized with COVID-19 (as of March 2020), though the drug was not formally approved through clinical trials.[46][47] From April

to June 2020, there was an emergency use authorization for their use in the United States,[48] and was used off label for potential treatment of the disease.[49] On 24 April 2020, citing the risk of "serious heart rhythm problems", the FDA posted a caution against using the drug for COVID-19 "outside of the hospital setting or a clinical trial".[40]

Their use was withdrawn as a possible treatment for COVID-19 infection when it proved to have no benefit for hospitalized patients with severe COVID-19 illness in the international Solidarity trial and UK RECOVERY Trial.[41][42] On 15 June 2020, the FDA revoked its emergency use authorization, stating that it was "no longer reasonable to believe" that the drug was effective against COVID-19 or that its benefits outweighed "known and potential risks".[43] In fall of 2020, the National Institutes of Health issued treatment guidelines recommending against the use of hydroxychloroquine for COVID-19 except as part of a clinical trial.[44]

In 2021, hydroxychloroquine was part of the recommended treatment for mild cases in India.[46]

In 2020, the speculative use of hydroxychloroquine for COVID-19 threatened its availability for people with established indications (malaria and autoimmune diseases).[48]

1.14 AIM OF PROJECT

The aims of this research work are

- (i) To synthesis novel complexes of chloroquine
- (ii) To characterize the resulting complexes using standard analytical techniques such as solubility, melting point, infrared and ultraviolet .

CHAPTER TWO

2.1 MATERIALS AND METHODS

2.1.1 APPARATUS

The following apparatus were used in the formation of the complexes and further analytical studies were carried out.

APPARATUS**MANUFACTURER**

Beakers

Pyrex scientific Ltd. England

Conical Flasks

Simax company Ltd. England

Capillary tube

Silber brand Ltd, England

Dessicator

Moncrief Scientific, England

Electrothermal melting point

Gallenkamp Ltd ,England

Round bottom flasks

Pyrex scientific Ltd, England

Hot plate with magnetic stirrer

Gallenkamp Ltd, England

Measuring cylinder

Technico scientific Ltd, England

Plastic condenser

Reflux condenser

Moramber (Mbc) Ltd

Test tube

Pyrex scientific Ltd, England

Thermometer

Uniscope scientific Ltd, England

Infrared

Duck V Scientific 500 infrared

Ultraviolet

Jenwoy 6405 UV spectrophotometer.

2.1.2 REAGENTS

The following reagents were collected from chemistry department, Kwara State Polytechnic.

Reagent	Manufacturer
Nickel Sulphate Hexahydrate	
Molecular Formular- $\text{NiSO}_4 \cdot 6\text{H}_2\text{O}$	J.T Barker Chemical C.O Philliburg
Molecular mass =262.71g/mol	
Copper Sulphate	
Molecular formula = CuSO_4 .	Eagle Scientific England
Molecular Mass= 159.60g/mol	
Iron Sulphate Heptahydrate	East Aglia, Chemicals Hadheigh
Molecular Formular= $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	
Molecular Mass =278.02g/mol.	
Ethanol	East Aglia, Chemicals Hadheigh
Methanol	East Aglia, Chemicals Hadheigh
Distilled water	

2.1.3 MATERIALS

Chloroquine

Molecular formular= $C_{18}H_{26}ClN_3$

Molecular Mass=319.872g/mol

Source Chemistry department, University of Ilorin.

2.2 EXPERIMENTAL PROCEDURES

All apparatus were cleaned thoroughly before use. The retort stand was set properly and a round bottom flask clamped to it. This was further positioned onto the hotplate with a magnetic stirrer. A reflux condenser was fixed into the round bottom flask containing the solution and clamped onto the retort stand. Water pipes were connected, one into the inlet and the other into the outlet with both connected to full bucket of water and an empty bucket respectively.

2.2.1 CHLOROQUINE Ni (ii) COMPLEX FORMATION

3.52g of chloroquine was weighed using a digital pocket scale and was then dissolved in 10cm³ of distilled water using a clean dried round bottom flask.

2.38g of NiSO₄.6H₂O was also weighed using a digital pocket scale and was then dissolved in 10cm³ of distilled water in a beaker.

The Nickel solution was added to the ibuprofen solution and shaken vigorously for it to mix well. The magnetic stirrer was placed in the mixture and it was refluxed for 1 hour after which the solution was allowed to cool and a light green color was observed. The resulting solution was filtered using a filter paper. The precipitate was then washed off with methanol.

The complex obtained was weighed and put in a container which was labeled CHLNi(ii) complex.

The weight of the complex obtained was 1.7g of methanol.

2.2.2 CHLOROQUINE Cu (ii) COMPLEX FORMATION

3.52g of chloroquine was weighed using a digital pocket scale and was then dissolved in 10cm³ of distilled water using a clean dried round bottom flask.

1.6g of CuSO₄ was also weighed and was then dissolved in 10cm³ of distilled water in a beaker. The magnetic stirrer was placed in the mixture and it was refluxed for 1 hour after which the solution was allowed to cool and a light green color was observed. The resulting solution was filtered using a filter paper. The precipitate was then washed off with methanol. The complex obtained was weighed and put in a container which was labeled IBCu(ii) complex. The copper solution was added to the chloroquine solution and shaken vigorously. The weight of the complex obtained was 3.5g.

2.2.3 CHLOROQUINE(Fe) FORMATION

3.52g of Chloroquine was weighed using a digital pocket scale and was then dissolved in 10cm³ of distilled water using a clean dried round bottom flask.

3.8g of FeSO₄ was also weighed and was then dissolved in 10cm³ of distilled water in a beaker. The magnetic stirrer was placed in the mixture and it was refluxed for 1 hour after which the solution was allowed to cool and a light brown color was

observed. The resulting solution was filtered using a filter paper. The precipitate was then washed off with methanol. The complex obtained was weighed and put in a container which was labeled CHLFe(ii) complex. The copper solution was added to the ibuprofen solution and shaken vigorously. The weight of the complex obtained was 3.2g.

2.3 CHARACTERIZATION METHODS

2.3.1 SolubilityTest.

The solubility of the complexes was determined in the following solvents. Ethanol, benzene, methanol, petroleum ether, acetone, distilled water and chloroform

2.3.2 MELTING POINT DETERMINATION

The melting point of the complexes were determined using Gallenkamp melting point apparatus, thermometer and capillary tubes

2.3.3 METHOD

One side of the capillary tubes was sealed and the samples were introduced through the open and to a depth of about 1cm of the tube. The thermometer and the capillary tubes were inserted in the hole provided in the apparatus. The

apparatus was switched on and the temperature at which the sample starts to melt and ends was taken as the melting point range of the sample. The melting point temperatures were taken for both the ligand and the complexes.

CHAPTER THREE

3.1 RESULTS AND DISCUSSION

The following are the results of the analysis conducted on some of the complexes formed

3.1.1 RESULTS OF SOLUBILITY TEST

Complex	Ethanol	Benzene	Methanol	Pet. Ether	Acetone	Distilled water	Chloroform
Chloroquine ligand	NS	NS	NS	NS	NS	S	NS
CHLNi(ii)	SS	NS	NS	NS	NS	SS	NS
CHLCu(ii)	SS	NS	NS	NS	NS	SS	NS
CHLFe(ii)	SS	NS	NS	NS	NS	SS	NS

Key S= soluble, SS=slightly soluble NS= not soluble

3.1.2 INTERPRETATION OF SOLUBILITY TEST

From the results obtained from the solubility test, it was evident that all complexes and the ligand were slightly soluble in methanol. Solubility of the ligand and the complexes were poor in ethanol, benzene, petroleum ether, acetone and chloroform.

3.2 RESULTS OF MELTING POINT TEST

COMPLEX	MELTING POINT RANGE
CHLOROQUINE LIGAND	190-192
CHLNi(ii) COMPLEX	158-160
CHLCu(ii) COMPLEX	160-162
CHLFe(ii) COMPLEX	211-213

3.2.1 INTERPRETATION OF MELTING POINT RANGE

From the results obtained, it showed that the melting point of the complexes were higher than those of the ligand. This shows that there was coordination.

Ligands/Complex	Wavelength(nm)	Absorbance	Transition Assignment
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Chloroquine	299.0	0.60	n- π^* transition
	260.0	1.756	π - π^* transition
CHLNi(ii)	278	2.916	n- π^* transition
	233.5	0.932	π - π^* transition
CHLCu(ii)	296	0.996	n- π^* transition
	251.5	0.638	π - π transition
CHLFe(ii)	284	2.328	n- π^* transition
	219	1.738	π - π transition

CHAPTER FOUR

4.1 CONCLUSION

Chloroquine formed stable complexes with Ni (ii), Cu (ii) and Fe (ii) ions. The solubility test and melting point determination showed that coordination has taken place. Further spectroscopic analysis like IR, UV and NMR is required in order to give a complete structure of the complexes.

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